

## Notes

INOSTAMYCINS B AND C,  
NEW POLYETHER ANTIBIOTICSHIDEHARU ODAI, KAZUTOSHI SHINDO\*,  
ATSUO ODAGAWA, JUNICHIRO MOCHIZUKI,  
MASA HAMADA and TOMIO TAKEUCHIPharmaceutical Research Laboratory,  
Kirin Brewery Co., Ltd.,  
3 Miyahara-cho, Takasaki-shi, Gunma 370-12, Japan  
\*Institute of Microbial Chemistry,  
3-14-23 Kamiosaki, Shinagawa-ku, Tokyo 141, Japan

(Received for publication March 4, 1994)

In the course of isolation of inostamycin (**1**), an inhibitor of phosphatidylinositol turnover, we found two related compounds, inostamycins B and C in the culture broth of *Streptomyces* sp. MH816-AF15.<sup>1,2)</sup> In this paper, we report the production, isolation, physico-chemical properties, structures and biological properties of inostamycin B (**2**) and C (**3**) (Fig. 1). These compounds showed antimicrobial activity against Gram-positive bacteria *in vitro*, but were not inhibitory to phosphatidylinositol turnover.

The producing strain was cultured in 500-ml Erlenmeyer flasks containing 100 ml of glycerol 2.0%, dextrin 2.0%, soy peptone 1.0%, yeast extract 0.3%, (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> 0.2%, and CaCO<sub>3</sub> 0.02%, the pH being adjusted to 7.4 before sterilization. The fermentation was carried out for 5 days at 30°C on a rotary shaker (180 rpm/minute). The culture broth (25 liters) was centrifuged and the mycelium cake

was extracted with acetone. The acetone extract was concentrated *in vacuo* and combined with the supernatant, then extracted with ethyl acetate. This extract was concentrated *in vacuo* and applied to a silica gel column which was developed with CHCl<sub>3</sub> and CHCl<sub>3</sub>-MeOH (100:1). Combined fraction containing inostamycins was further purified by centrifugal partition chromatography (Sanki Engineering Limited, model LLN, solvent system: acetonitrile-hexane) to afford pure **1** (965 mg), **2** (29.8 mg) and **3** (38.3 mg).

Physico-chemical properties of **2** and **3** are summarized in Table 1. <sup>13</sup>C NMR chemical shifts of these antibiotics are shown in Table 2.

The molecular ion peak of **2** was observed in the FD-MS spectrum at *m/z* value 686, which is less than **1** by 14 mass units. Comparison of <sup>1</sup>H NMR spectra of **1** and **2** showed that one triplet methyl signal (38-H, δ<sub>H</sub> 0.93) in **1** was replaced by a doublet methyl signal (37-H, δ<sub>H</sub> 1.28) in **2**. Furthermore, a long range coupling was observed from the doublet methyl signal to the carboxyl group (C-1, δ<sub>C</sub> 179.5) in the HMBC spectrum of **2**. Therefore, it was concluded that the ethyl group at C-2 in **1** was replaced to a methyl group in **2** (Fig. 1). Closely related compounds have recently been reported.<sup>3)</sup>

The FD-MS spectrum of **3** gave a dehydration peak at *m/z* value 638. In the <sup>13</sup>C NMR spectra of **3**, one carboxyl group (C-1, δ<sub>C</sub> 181.05) and one methine signal (C-2, δ<sub>C</sub> 55.90) present in **1** was replaced by one methylene signal (C-2, δ<sub>C</sub> 41.6). Thus, **3** was confirmed to be a decarboxyl compound of **1** (Fig. 1). Similar decarboxylation products were reported by KOENUMA *et al.*<sup>4)</sup>

Fig. 1. The structures of inostamycin, inostamycin B and C.

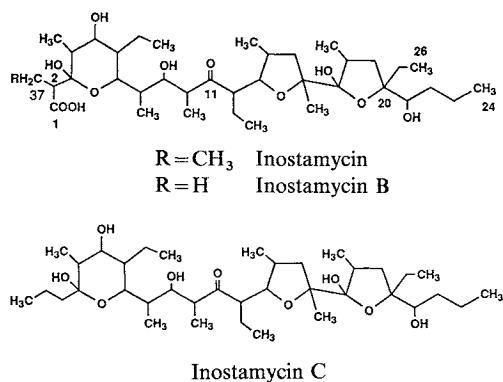


Table 1. Physico-chemical properties of inostamycins B and C.

	Inostamycin B	Inostamycin C
MP	82~83°C	148~150°C
[α] <sub>D</sub> <sup>26</sup>	+3.0	+2.6
(c 0.5, CHCl <sub>3</sub> )		
Molecular formula	C <sub>37</sub> H <sub>66</sub> O <sub>11</sub>	C <sub>37</sub> H <sub>68</sub> O <sub>9</sub>
FD-MS	686 (M) <sup>+</sup>	638 (M-H <sub>2</sub> O) <sup>+</sup>
Analysis	Calcd:	Found: Calcd: Found:
C	64.68	63.88 67.63 68.02
H	9.69	9.36 10.44 10.13
Rf <sup>a</sup> in silica gel	0.41	0.66
TLC		

<sup>a</sup> CHCl<sub>3</sub>-MeOH (20:1).

Table 2.  $^{13}\text{C}$  chemical shift assignment of inostamycins in  $\text{CDCl}_3$ .

	Inostamycin <sup>a</sup>	Inostamycin B	Inostamycin C		Inostamycin <sup>a</sup>	Inostamycin B	Inostamycin C
1 s	181.05	179.5		20 s	87.26	88.7	88.3
2-CH d	55.90	43.3		21 d	69.97	73.8	70.9
2-CH <sub>2</sub> t			41.6	22 t	34.82	33.6	34.0
3 s	100.69	100.6	100.2	23 t	20.28	19.4	19.4
4 d	38.22	38.6	38.9	24 q	14.40	14.1	14.1
5 d	71.17	71.6	71.5	25 t	31.11	30.8	30.4
6 d	37.57	36.8	36.6	26 q	7.18	7.5	7.4
7 d	74.75	75.8	75.1	27 q	14.74	12.1	13.7
8 d	32.30	33.1	32.8	28 q	23.98	24.2	24.3
9 d	76.50	78.0	78.1	29 q	15.56	14.2	15.0
10 d	47.48	48.2	47.5	30 t	14.99	14.6	16.5
11 s	214.96	210.8	213.3	31 q	12.54	12.7	12.5
12 d	55.16	54.1	55.2	32 q	12.84	13.5	13.7
13 d	83.62	82.3	84.2	33 q	5.28	4.6	4.7
14 d	34.73	33.9	35.2	34 t	18.41	20.0	18.9
15 t	42.68	42.1	42.0	35 q	10.85	11.2	10.8
16 s	86.29	82.9	83.9	36 q	13.17	13.1	13.7
17 s	108.30	107.6	107.4	37-CH <sub>2</sub> t	20.08		15.9
18 d	38.42	38.9	38.0	37-CH <sub>3</sub> q		12.8	
19 t	37.57	35.6	36.7	38 q	12.37		13.8

<sup>a</sup> Cited from the data by IMORO *et al.*<sup>1)</sup>

Table 3. Antimicrobial activities of inostamycins.

Test organism	MIC ( $\mu\text{g/ml}$ )		
	Inostamycin	Inostamycin B	Inostamycin C
<i>Staphylococcus aureus</i> FDA 209P	0.78	1.56	3.12
<i>S. aureus</i> Smith	0.78	3.12	100
<i>S. aureus</i> MS9610	0.78	3.12	100
<i>S. aureus</i> No. 5	0.78	3.12	100
<i>S. aureus</i> No. 17	0.78	6.25	> 100
<i>Micrococcus luteus</i> FDA 16	0.78	3.12	12.5
<i>M. luteus</i> IFO 3333	0.78	3.12	12.5
<i>M. luteus</i> PCI 1001	0.78	3.12	> 100
<i>Bacillus anthracis</i>	0.78	1.56	6.25
<i>B. subtilis</i> NRRL B-558	0.78	> 100	> 100
<i>B. subtilis</i> PCI 219	0.78	6.25	> 100
<i>B. subtilis</i> ATCC 10702	0.78	1.56	6.25
<i>Corynebacterium bovis</i> 1810	0.78	3.12	6.25
<i>Mycobacterium smegmatis</i> ATCC 607	1.56	> 100	> 100
<i>Escherichia coli</i> NIHJ	> 100	> 100	> 100
<i>E. coli</i> K-12	> 100	> 100	> 100
<i>Shigella dysenteriae</i> JS 11910	> 100	> 100	> 100
<i>Salmonella typhi</i> T-63	> 100	> 100	> 100
<i>Pseudomonas aeruginosa</i> A3	100	> 100	> 100
<i>Klebsiella pneumoniae</i> PCT 602	> 100	> 100	> 100
<i>Candida albicans</i> 3147	100	> 100	> 100

Mueller-Hinton agar, 37°C.

Inostamycins showed antimicrobial activities against Gram-positive bacteria. The results are given in Table 3. The antimicrobial activities of 2 and 3 are relatively reduced compared to those of 1.

Inostamycins have cytotoxic activity against *src*-NIH-3T3 cells. IC<sub>50</sub> values of 1, 2 and 3 were 0.07, 0.5 and 0.5  $\mu\text{g/ml}$ , respectively.

## References

- 1) IMOTO, M.; K. UMEZAWA, Y. TAKAHASHI, H. NAGANAWA, Y. IITAKA, H. NAKAMURA, Y. KOIZUMI, Y. SASAKI, M. HAMADA, T. SAWA & T. TAKEUCHI: Isolation and structure determination of inostamycin, a novel inhibitor of phosphatidylinositol turnover. *J. Natl. Prod.* 53: 825~829, 1990
- 2) IMOTO, M.; Y. TANIGUCHI & K. UMEZAWA: Inhibition of CDP-DG: inositol transferase by inostamycin. *J. Biochem.* 112: 299~302, 1992
- 3) WESTLEY, J. W.; C-M. LIU, J. F. BLOUNT, L. TODARO, L. H. SELLO & N. TROUPE: Isolation and characterization of four polyether antibiotics, X-14889A, B, C and D, closely related to lysocellin and the ferensimycins. *J. Antibiotics* 46: 280~286, 1993
- 4) KOENUMA, M & N. ŌTAKE: Studies on the ionophorou antibiotics. XI The artifacts and the degradation products of lysocellin. *J. Antibiotics* 30: 819~828, 1977